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Tetrahedron Letters

journal homepage: www.elsevier.com/locate/tetlet

Synthesis of *ent*-gabosine E from **D**-mannose by intramolecular nitrone–olefin cycloaddition

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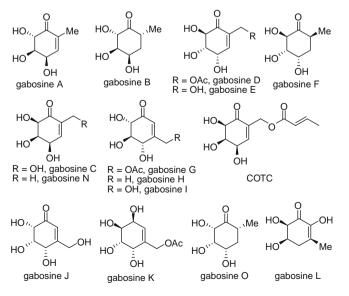
ARTICLE INFO	A B S T R A C T
Article history: Received 21 August 2009 Revised 11 September 2009 Accepted 25 September 2009 Available online 1 October 2009	Intramolecular nitrone-olefin cycloaddition in a mannose template followed by N-O cleavage, quatern- ization of the resulting amine, and finally oxidative elimination of the amino group affords, after depro- tection, <i>ent</i> -gabosine E in enantiomerically pure form. © 2009 Elsevier Ltd. All rights reserved.

The name 'gabosines' has been given to a group of 14 carba-sugars isolated from *Streptomyces* strains.¹ The majority are diastereoisomeric 4,5,6-trihydroxy-2-cyclohexenones with an additional methyl, hydroxymethyl, or acetoxymethyl group at the 2- or 3positions (Fig. 1), and exhibit a variety of biological activities such as antiprotozoal activity, DNA binding properties, and enzyme inhibition. Related natural products include the glyoxylase I inhibitor COTC² (Fig. 1) isolated from *Streptomyces filipensis*, the plant growth factor streptol,^{1c} its diastereoisomer MK7607³ isolated from *Curvularia eragrostidis* which possesses herbicidal activity, the antibiotic rancinamycins,^{4a} and numerous structurally related natural products.⁴ Furthermore, gabosine-like compounds are constituents of complex entities such as acarbose and/or have been used as intermediates for the synthesis of biologically interesting compounds.

As a consequence, a number of synthetic approaches toward gabosines and related compounds have been reported in the literature,⁵ usually starting from carbohydrates or quinic acid. Despite the existence of these synthetic methods for converting sugars into gabosines, a general route starting from a monosaccharide leading to a 4,5,6-trihydroxy-2-hydroxymethyl-2-cyclohexenone with transfer of chirality to the products and allowing the synthesis of any of its diastereoisomers, is needed. Such general methods exist only for gabosines with a 3-hydroxymethyl⁵ⁱ or 2-methyl^{5k} substituent. Whilst investigating a general synthetic scheme to gabosines with a 3-hydroxymethyl substituent, we considered that our recent work⁶ on the synthesis of chiral cyclopentenones might be useful and applicable in this case.

Starting from a hexose, a carbocyclic ring with a hydroxymethyl group could be prepared by an intramolecular nitrone–olefin cycloaddition and further manipulation of the cycloadduct (Scheme 1). The installation of an enone group in protected gabosine **1** could be accomplished by quaternization and subsequent

oxidative elimination of hydroxyamine **2**, which could be obtained by N–O bond cleavage in bicyclic isoxazolidine **3**. The latter is the intramolecular cycloadduct of the nitrone derived from the sugar template **4**, which in turn is easily prepared from a hexose. Thus, following this route and assuming that no epimerization would occur during the reaction sequence, the absolute configurations of the C-2, C-3, and C-4 chiral centers of the starting sugar should be transferred to the C-4, C-5, and C-6 positions of the final gabosine. Importantly, this sequence gives us the opportunity to selectively protect the three secondary hydroxy groups, allowing oxidation of the fourth hydroxyl as necessary. It is also worth noting that the stereoselectivity of the cycloaddition reaction is of less









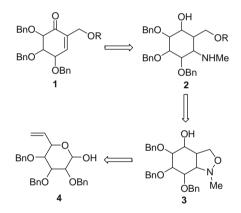
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importance, since the newly formed stereocenters are subsequently destroyed.

D-Mannose was the starting material of choice, which would result in ent-gabosine E by transfer of its chirality to the final product, applying the above-mentioned methodology. Using the standard protection-deprotection manipulations, commercially available methyl α -D-mannopyranoside **5** was converted into the known compound **6** according to the literature⁷ (Scheme 2). Next, the free primary hydroxy group was subjected to Swern oxidation and Wittig reaction to give the known template **7**.⁸ Acetal hydrolysis was achieved by the treatment of 7 with H₂SO₄ and Ac₂O followed by hydrolysis of the intermediate acetate with EtONa in EtOH to give 8 in excellent yield. This intermediate was able to undergo the intramolecular nitrone cycloaddition upon condensation with MeNHOH.⁹ After some experimentation, we found that the treatment of MeNHOH HCl with EtONa/EtOH for 30 min, followed by addition to 8 gave the respective nitrone, which without isolation, afforded upon stirring for 24 h, the two cycloaddition products 9 and 10 in 80% combined yield and in a 2:1 ratio. It is interesting to note that when pyridine was used as the solvent and base in the reaction of 8 with MeNHOH HCl, partial epimerization of the C-2 (sugar numbering) stereocenter took place. The two products 9 and 10 were separated by chromatography and their structures were elucidated by COSY and NOE experiments.^{10,11}

The N–O bond of the major cycloadduct **9** was then cleaved using a standard procedure (Zn, AcOH) and the primary hydroxy



Scheme 1. Retrosynthetic analysis of gabosine.

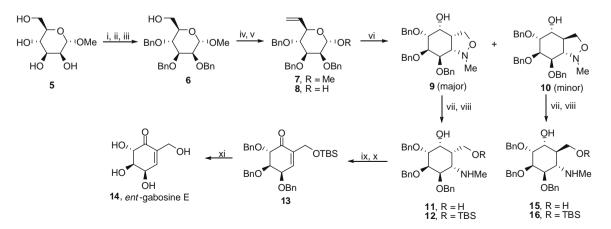
group was selectively protected using TBSOTf to give compound **12** in 77% overall yield (two steps). Quaternization of the amino group of **12** followed by oxidative elimination led to protected *ent*-gabosine E **13**. This method has been used previously by us,⁶ and others¹² for the regioselective formation of an enone functionality. Finally, the protecting groups were removed from **13** with BBr₃ and the expected *ent*-gabosine E **14** was obtained in 85% yield with spectroscopic data in good agreement with those reported for gabosine E.^{1a,13}

Unfortunately, the minor cycloadduct **10** did not give **14** when we applied the same reaction sequence. Attempted oxidation of the free secondary hydroxy group after N–O bond scission to **15**, protection of the primary hydroxy group and quaternization of the amine did not give *ent*-gabosine, evidently because the H and Me₃N groups are not *anti*-disposed for elimination, a fact that confirmed the assigned structure of **10**.

In summary, we have prepared enantiomerically pure *ent*-gabosine E in 11 steps and 12% overall yield starting from commercially available methyl α -p-mannopyranoside, using generally applicable reactions and procedures. In addition, interesting new aminocyclitols, such as **11** and **15**, are prepared as intermediates. Similarly, starting from other hexoses, diasteroisomers of **14** may be prepared and work in this direction is in progress.

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Scheme 2. Reagents and conditions: (i) TrCl, pyridine, 20 °C, 24 h, 84%; (ii) BnCl, NaH, DMF, 20 °C, 12 h, 90%; (iii) H₂SO₄, MeOH (abs), 20 °C, 1 h, 84%; (iv) (COCl)₂, DMSO, Et₃N, DCM, -60 °C then Ph₃P*CH₃Br⁻, *n*-BuLi 1.6 M in hexanes, THF, -70 °C, 1 h, 72%; (v) H₂SO₄, Ac₂O, 0 °C, 10 min, then EtONa, EtOH, 0 °C, 10 min, 91%; (vi) MeNHOH HCl, EtONa, EtOH, then 20 °C, 24 h, 80%; (vii) Zn, AcOH, reflux, 1 h; (viii) TBSOTf, 2,6-lutidine, CH₂Cl₂, -78 °C, 45 min, 77% for **12**, 80% for **16**; (ix) excess Mel, K₂CO₃, THF, 24 h; (x) Dess-Martin periodinane, CH₂Cl₂, 20 °C, 30 min, 80% from **12**; and (xi) BBr₃, CH₂Cl₂, -78 °C, 45 min, 85%.

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- 10. The proton signal assignment of diastereoisomers 9 and 10 was made by H,H-COSY experiments. Significant NOE enhancements were observed between the two bridged protons (3a-H and 7a-H) as well as between 3a-H and 4-H in compound 9, whereas no considerable enhancement was shown between 7-H and 7a-H. On the other hand, in compound 10, an NOE enhancement was observed between 7-H and 3a-H, but no significant enhancements between the two bridged protons (3a-H and 7a-H) as well as between the couples 3a-H, 4-H and 7-H, 7a-H were evident.
- 11. Selected data. *Compound* **9**: oil, $[\alpha]_D 54.2$ (*c* 1.0, CHCl₃); IR (neat film) 3500 (br) cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 2.66 (s, 3H, Me), 2.70 (m, 1H, 6-H), 3.15 (m, 2H, 1-H), 3.79 (dd, *J* = 5.2, 4.0 Hz, 1H), 3.85 (dd, *J* = 6.4, 2.8 Hz, 1H), 3.89 (dd, *J* = 5.2, 2.8 Hz, 1H), 4.06 (t, *J* = 8.0 Hz, 1H), 4.11 (dd, *J* = 7.2, 6.4 Hz, 1H), 4.20 (br s, 1H), 4.48 (d, *J* = 12.0 Hz, 1H), 4.52 (d, *J* = 12.0 Hz, 1H), 4.62 (d, *J* = 12.0 Hz, 1H), 4.64 (d, *J* = 12.0 Hz, 1H), 4.62 (d, *J* = 12.0 Hz, 1H), 4.64 (d, *J* = 12.0 Hz, 1H), 4.672 (d, *J* = 12.0 Hz, 1H), 4.62 (d, *J* = 12.0 Hz, 1H), 4.68 (d, *J* = 12.0 Hz, 1H), 4.72 (d, *J* = 12.0 Hz, 1H), 7.25-7.40 (m, 15H); ¹³C NMR (CDCl₃, 75 MHz) δ 44.6, 44.8, 66.8, 67.2, 68.3, 72.6, 73.2 (two peaks), 75.4, 76.1, 79.5, 127.6, 127.7 (two peaks), 127.8, 127.9, 128.3, 128.4, 128.5, 137.8, 138.5, 138.7; HRMS *m*/z 476.2440 [C₂₉H₃₄NO₅ (M+H)⁺ requires 476.2431]. *Compound* 10: oil, $[\alpha]_D 48.7$ (*c* 0.9, CHCl₃); IR (neat film) 3500 (br) cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 2.10 (br s, 1H), 2.69 (m, 1H), 2.83 (t, *J* = 10.8 Hz, 1H), 2.85 (s, 3H), 3.69 (t, *J* = 3.2 Hz, 1H), 3.83 (m, 3H, 2-H),

3.94 (dd, J = 10.8, 3.2 Hz, 1H), 4.07 (t, J = 6.8 Hz, 1H), 4.33 (d, J = 12.0 Hz, 1H), 4.36 (d, *J* = 12.0 Hz, 1H), 4.42 (d, *J* = 12.0 Hz, 1H), 4.44 (d, *J* = 12.0 Hz, 1H), 4.48 (d, *J* = 12.0 Hz, 1H), 4.73 (d, *J* = 12.0 Hz, 1H), 7.25–7.40 (m, 15H); $^{13}{\rm C}$ NMR (CDCl₃, 75 MHz) δ 47.5, 49.4, 67.7, 68.2, 69.5, 71.9, 73.2, 73.7, 74.8, 79.6, 80.4, 127.7, 127.9, 128.1, 128.4, 128.6, 137.7, 138.0, 138.1; HRMS m/z 476.2438 [C₂₉H₃₄NO₅ (M+H)⁺ requires 476.2431]. Compound **12**: oil, $[\alpha]_D - 12.7$ (c 1.5, CHCl₃); IR (neat film) 3325 cm⁻¹; ¹H NMR (C₆D₆, 400 MHz, some peaks are broad due to conformer interconversion) δ 0.11 (s, 3H), 0.12 (s, 3H), 1.01 (s, 9H), 2.17 (s, 3H), 2.34 (br m, 1H, 6-H), 3.07 (br m, 1H), 3.95 (dd, J = 10.4, 3.2 Hz, 1H), 4.04 (m, 4H), 4.23 (br, 1H), 4.49 (d, J = 12.0 Hz, 1H), 4.60 (d, J = 12.0 Hz, 1H), 4.62 (d, J = 12.0 Hz, 1H), 4.64 (d, J = 12.0 Hz, 1H), 4.60 (d, J = 12.0 Hz, 1H), 4.75 (d, J = 12.0 Hz, 1H), 4.81 (d, J = 12.0 Hz, 1H), 7.05–7.40 (m, 15H); 13 C NMR (CDCl₃, 100 MHz) δ –5.4, 18.2, 25.9, 34.7, 39.9, 58.7, 61.3, 69.6, 72.5, 72.9, 73.3, 75.9, 76.4, 80.2, 127.4, 127.5, 127.8, 128.3, 138.7, 138.8, 139.1; HRMS *m/z* 592.3469 [C₃₅H₅₀NO₅Si (M+H)⁺ requires 592,3453]. Compound **13**: oil, $[a]_D$ – 91.1 (c 1.2, CHCl₃); IR (neat film) 1680 cm⁻¹; ¹H NMR (CDCl₃, 400 MHz) δ 0.08 (s, 3H), 0.09 (s, 3H), 0.93 (s, 9H), 3.96 (dd, J = 8.0, 3.2 Hz, 1H), 4.36 (m, 3H), 4.44 (br s, 1H), 4.69 (d, J = 11.2 Hz, 1H), 4.72 (d, J = 12.0 Hz, 2H), 4.79 (d, J = 12.0 Hz, 1H), 4.81 (d, J = 12.0 Hz, 1H), 4.91 (d, J = 11.2 Hz, 1H), 6.88 (d, J = 4.8 Hz, 1H), 7.25–7.40 (m, 15H); ¹³C NMR (CDCl₃, 100 MHz) δ –5.5, –5.45, 18.3, 25.9, 59.5, 72.2, 72.5, 73.2, 74.0, 78.8, 80.3, 127.7 (two peaks), 127.8, 128.0, 128.3 (two peaks), 128.4, 137.8, 138.1, 138.2, 138.6, 138.9, 196.4; HRMS m/z 559.2882 [C₃₄H₄₃O₅Si (M+H)⁺ requires 559.2874]. Compound 16: oil; IR (neat film) 3325 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 0.11 (s, 3H), 0.12 (s, 3H), 0.90 (s, 9H), 2.05 (m, 1H), 2.60 (s, 3H), 3.13 (t, J = 10.8 Hz, 1H), 3.76 (m, 3H), 3.87 (dd, J = 10.5, 7.5 Hz, 1H), 4.03 (d, J = 8.7 Hz, 1H), 4.12 (dd, J = 10.8, 4.5 Hz, 1H), 4.27 (d, J = 11.1 Hz, 1H), 4.43 (d, J = 11.1 Hz, 1H), 4.44 (d, J = 12.0 Hz, 2H), 4.58 (d, J = 12.0 Hz, 1H), 4.64 (d, J = 12.0 Hz, 1H), 7.20– 7.40 (m, 15H); ¹³C NMR (CDCl₃, 100 MHz) δ –5.8, –5.7, 18.1, 25.8, 33.9, 42.0, 60.1, 64.5, 67.6, 71.5, 71.9, 73.2, 73.9, 76.5, 77.8, 127.8, 128.0 (two peaks), 128.1, 128.2, 128.4 (two peaks), 128.5, 128.8, 137.4, 137.7, 137.8; HRMS m/z 592.3460 [C₃₅H₅₀NO₅Si (M+H)⁺ requires 592.3453].

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- 13. For ent-gabosine E **14**, $[\alpha]_D^{00} 145.7$ (*c* 0.35, MeOH) [for gabosine E lit.^{1a} $[\alpha]_D$ +148 (*c* 0.85, MeOH) and +152 (*c* 1.0, H₂O)].